

## **Immunofluorescence microscopy sample preparations and stainings**

- 1.□ infect cells as described for invasion assay
- 2.□ fix cells on coverslips for 25min @ RT with 4% EM grade paraformaldehyde
- 3.□ wash well with PBS
- 4.□ block 15 min with serum or BSA i.e. 1% normal goat serum if using secondary from goat
- 5.□ add primary antibody diluted in PBS + %BSA + 0.1% saponin +1% normal goat serum (1/ 200 anti-FLAG monoclonal made in mouse)
- 6.□ incubate in humid dark chamber 1-3 @ RT or as long as O/N @ 4°C
- 7.□ wash with PBS
- 8.□ incubate with diluted secondary antibody (dil in same as primary) ie 1/200 goat anti-mouse-TRITC
- 9.□ wash w/ PBS
- 10.□ can stain plasma membrane with DiD @ RT for 15 min  
2µl stock+ 250 µl DMSO + 1.648 ml PBS
- 11.□ wash and mount with anti-bleach media
- 12.□ seal all edges of coverslip with nail polish