Preparing Paraffin Tissue Sections and Stainings
From Denise Monack

1. cut small blocks of tissue approx. 1cm^2 x 0.4 cm
2. place in buffered formalin
3. incubate 2 h to overnight
4. paraffin embedding done by Lisa Raskin as well as sectioning
5. place in 60°C oven for 30 min
6. dewax in xylene bath, 2x 5 min
7. 96% ethanol, 2x 5 min
8. 90% ethanol
9. 80% ethanol
10. double distilled water, 2x 5 min
11. nuclei of tissue sections stripped from proteins by incubation with 20 m□□□
12. wash in DDW for 2 min, four times
13. rinse in PBS, 2x 5 min
14. dry area around tissue, mark with Pap pen and then overlay tissue with TUNEL reaction (I use Boehringer Mannheim kit)
15. incubate in humidified chamber for 60 min @ 37°C
16. rinse w/ PBS, 2x 5 min
17. add primary antibodies diluted in PBS, 3% BSA, 0.1% saponin
18. incubate 1 h @ room temp
19. wash in PBS, 4x 5 min
20. incubate in secondary antibodies diluted in same sol as before and incubate for 1 h @ room temp
21. wash in PBS, 4x 5 min
22. mount with antiquench (Vectashield) and overlay with large coverslip